



Cite this article: Finnegan S, Rasmussen CMØ, Harper DAT. 2016 Biogeographic and bathymetric determinants of brachiopod extinction and survival during the Late Ordovician mass extinction. *Proc. R. Soc. B* **283**: 20160007.
<http://dx.doi.org/10.1098/rspb.2016.0007>

Received: 2 January 2016

Accepted: 4 April 2016

Subject Areas:

palaeontology, evolution, ecology

Keywords:

extinction, climate change, biogeography, Ordovician, brachiopoda

Author for correspondence:

Seth Finnegan

e-mail: sethf@berkeley.edu

Electronic supplementary material is available at <http://dx.doi.org/10.1098/rspb.2016.0007> or via <http://rspb.royalsocietypublishing.org>.

Biogeographic and bathymetric determinants of brachiopod extinction and survival during the Late Ordovician mass extinction

Seth Finnegan¹, Christian M. Ø. Rasmussen^{2,3,5} and David A. T. Harper^{4,5}

¹Department of Integrative Biology, University of California, Berkeley, CA, USA

²Natural History Museum of Denmark, and ³Center for Macroecology, Evolution and Climate, University of Copenhagen, Øster Voldgade 5–7, 1350 Copenhagen, Denmark

⁴Palaeoecosystems Group, Department of Earth Sciences, Durham University, Durham DH1 3LE, UK

⁵Department of Geology, University of Lund, Lund, Sweden

The Late Ordovician mass extinction (LOME) coincided with dramatic climate changes, but there are numerous ways in which these changes could have driven marine extinctions. We use a palaeobiogeographic database of rhynchonelliform brachiopods to examine the selectivity of Late Ordovician–Early Silurian genus extinctions and evaluate which extinction drivers are best supported by the data. The first (latest Katian) pulse of the LOME preferentially affected genera restricted to deeper waters or to relatively narrow (less than 35°) palaeolatitudinal ranges. This pattern is only observed in the latest Katian, suggesting that it reflects drivers unique to this interval. Extinction of exclusively deeper-water genera implies that changes in water mass properties such as dissolved oxygen content played an important role. Extinction of genera with narrow latitudinal ranges suggests that interactions between shifting climate zones and palaeobiogeography may also have been important. We test the latter hypothesis by estimating whether each genus would have been able to track habitats within its thermal tolerance range during the greenhouse–icehouse climate transition. Models including these estimates are favoured over alternative models. We argue that the LOME, long regarded as non-selective, is highly selective along biogeographic and bathymetric axes that are not closely correlated with taxonomic identity.

1. Introduction

The Late Ordovician mass extinction (LOME) was one of the largest extinctions of the past 500 million years [1,2], involving the extinction of almost half of marine invertebrate genera and an estimated approximately 85% of species [3]. Major extinction pulses occurred at the boundary between the Katian and Hirnantian stages, and in the mid-Hirnantian (at the base of the *Metabolograptus persculptus* graptolite Biozone) [3,4]. Both these boundaries are associated with major climatic and oceanographic transitions: the former with global cooling, expansion of south polar ice sheets and falling sea levels, and the latter with warming, melting of ice sheets and continental flooding [3–9].

Because of the apparent correspondence between climatic transitions and extinction pulses, the LOME has long been recognized as a climatically driven event [3–8,10]. However, substantial uncertainty remains regarding the *mechanisms* of extinction—there are many ways in which climatic transitions and associated oceanographic changes could potentially lead to elevated extinctions, particularly through habitat loss [5,8,11]. Previous studies of the LOME have implicated two major drivers of extinction: (i) loss of species inhabiting shallow cratonic seaways that drained as Gondwanan glaciers grew; and

(ii) loss of species with narrow and/or relatively warm thermal tolerance ranges as the polar front advanced and the latitudinal temperature gradient steepened [12,13]. Late Ordovician–Early Silurian climatic events seem also to have been associated with major changes in oceanographic circulation, productivity and oxygenation of outer shelf and slope settings [6,14]. Changes in oxygenation, in particular, have been suggested as a major additional agent of extinction, particularly for taxa with planktonic life stages [15–17]. Finally, the fact that the Hirnantian stage is represented by hiatuses, lacunae or major facies shifts in many regions [6,13,18–20] and the preponderance of ‘Lazarus’ genera that are missing from the Hirnantian and earliest Silurian records, but later reappear [21], suggest that some proportion of apparent losses during the LOME may be artefacts of record failure.

A recent analysis of sequence stratigraphic architecture in Upper Ordovician sections in Gondwana and tropical Laurentia suggested that the first pulse of the LOME does not in fact coincide with cooling and sea level fall but rather with the first major interglacial episode of the Hirnantian stage [19]. This interpretation, if supported, would require a radical reconsideration of the nature of the LOME and the lessons that can be drawn from it regarding general relationships between climate change and extinction in the fossil record.

A comprehensive analysis of latest Ordovician extinction selectivity patterns and their context in broader Late Ordovician–Early Silurian selectivity patterns is therefore needed to evaluate relative support for different causal models of the LOME. Here we use a large and taxonomically standardized occurrence database [22–24] to examine Late Ordovician–Early Silurian extinction patterns within one of the most diverse and well-preserved Palaeozoic clades, the rhynchonelliform brachiopods.

2. Material and methods

See the electronic supplementary material for additional details on data and analyses.

(a) Database

Our dataset was compiled from the literature and from ongoing research programmes in Durham and Copenhagen. Local stratigraphic ranges of rhynchonelliform brachiopods were recorded to the species level where possible. In the literature, though, only generic lists are often published. Thus, we omitted species-level data for this study and focused on genera. Stratigraphically, all data have been correlated with the British stage system using Bergström *et al.* [25] in order to standardize correlation. For this study, we have translated the regional stages into global units, again using Bergström *et al.* [25]. Taxonomic revisions dealing with specific genera have been assessed and their recommendations implemented wherever possible to avoid synonyms. Brachiopod species, and genera exhibit strong depth preferences and benthic assemblage (BA) zones have long been used as depth indicators in the early Palaeozoic [26–28]. Each genus in our dataset was assigned to a range of BAs based on information in the literature, often with reference to associated fauna or lithology. In cases for which literature data were not available, assignments were based on the authors’ experience with faunal and facies associations in the field. Further details regarding the database are available in previous publications [22–24].

(b) Quantifying extinction and risk predictors

All data manipulations and analyses were carried out in the R programming environment [29]. For each of 23 Late Ordovician–Early Silurian timeslices resolvable in our database (electronic supplementary material), we tallied all genera that were sampled in at least one locality (i.e. the timeslice falls within the local stratigraphic range of the genus at least one locality). To avoid edge effects, we excluded the Sandbian (early Late Ordovician) and Telychian (late Early Silurian) intervals. In the remaining 19 timeslices, the number of extant genera ranges from 121 (earliest Rhuddanian) to 238 (earliest Katian). Within each timeslice, we count genera as survivors if they are known to occur in younger timeslices and extinctions if they are not. We further divided survivors into genera that are sampled in at least one region in the immediately succeeding timeslice and ‘Lazarus’ genera [21,30] that disappear from the record for at least one timeslice but subsequently reappear. In keeping with common palaeobiological practice, we analysed geographical and stratigraphic ranges at the genus level, because species are inconsistently identified in the literature. If the determinants of extinction risk differ between the species and genus levels, then species-poor genera are expected to be at higher risk than relatively speciose genera. To account for such differences, we tallied the number of named species assigned to each genus during each timeslice (species richness). Genera lacking named species (e.g. all occurrences in that timeslice were recorded as ‘sp.’) were assigned a minimal value of 1.

Within each timeslice, we tabulated several aspects of geographical distribution for each genus. These included great circle distance (the maximum distance between any two localities occupied by a genus during a given timeslice; genera occurring at only a single locality were assigned a value of 1), grid cell occupancy (the number of 10° latitude by 30° longitude grid cells that contain at least one locality occupied by a genus during a given timeslice) and number of palaeocontinents (the number of palaeocontinents and terranes occupied by a genus during a given timeslice). We quantified latitudinal distribution by tabulating the minimum and maximum absolute palaeolatitudes at which each genus occurs in each timeslice and its absolute palaeolatitudinal range (e.g. from 0° to 90°). We quantified bathymetric distribution by tabulating the minimum and maximum depth (as measured by BA membership) and the depth range of each genus. To evaluate whether genera entirely or largely confined to cratonic seaways were harder hit than open-shelf genera we tabulated % cratonic localities (the percentage of all localities occupied by each genus in each timeslice that were located in cratonic seaways).

The sharp decline in the amount of preserved sedimentary rock between the Katian and the Hirnantian [13] raises the possibility that some of the apparent extinction during the first pulse of the LOME reflects record failure rather than reflecting true extinction. No existing database captures the global distribution of sedimentary rock with spatio-temporal resolution sufficient to compare directly with our database. Instead, in each timeslice, we categorized each locality in our database as continuous if at least one brachiopod genus occurred at that locality in the immediately succeeding timeslice, and discontinuous if no brachiopod genera are documented from that locality in the succeeding timeslice. A given locality may appear to be discontinuous in the immediately succeeding timeslice for several reasons, including (i) absence of sedimentary rock owing to non-deposition and/or subsequent erosion, (ii) lack of exposure, (iii) lack of expert collecting effort and (iv) lack of appropriate palaeoenvironments (e.g. rocks are terrestrial, or anoxic marine black shales), but all of these scenarios represent failures of preservation and/or collection that have the effect of truncating true stratigraphic ranges. To determine whether such truncations were an important predictor of apparent extinction risk, we

tabulated % discontinuous localities (the percentage of all localities occupied by a genus during a given timeslice that have no records in the succeeding timeslice).

For the latest Katian timeslice (Katian 5), we undertook a further set of analyses to evaluate whether extinction risk was influenced by interactions between biogeographic distribution and shifting climate zones (see the electronic supplementary material). Using published climate simulations [31], palaeogeographic reconstructions [32,33] and observed palaeogeographic distributions, we estimated minimal thermal tolerance ranges for all genera extant during Katian 5. We then determined whether each regional population in a given genus would have been able to access habitat within this thermal range by shifting its range equatorward during the greenhouse–icehouse transition (electronic supplementary material, figure S1). Genera with at least one population that should have been able to remain within its thermal tolerance range without crossing an ocean basin were counted as predicted survivors. Those for which no suitable habit would have been available without dispersing across open oceans were counted as predicted extinctions. For comparison, we also made predictions based on the opposite scenario: an assumed icehouse state in the latest Katian transitioning to a Hirnantian greenhouse state.

(c) Modelling relationships between predictors and extinction risk

We examined relationships between the predictors described above and extinction patterns in the nine timeslices (out of 19 total) that had at least 10 observed genus extinctions. We used generalized boosted regression models (GBMs) [34,35] to determine which predictors were most strongly associated with extinction risk in each interval and the marginal effect (the effect with all other variables held constant) of varying each predictor on extinction risk. GBM models are useful for delineating extinction selectivity patterns throughout the study interval, but ensemble-based models of this kind cannot easily be compared in a maximum-likelihood framework. To make explicit models comparisons for the Katian 5 timeslice, we constructed multivariate binomial logistic regression models [36] using the subset of predictors that were included in 75% or more of the top-ranked models (those that had AICc scores within four points of the ‘best’ model). These predictors were palaeolatitude range, minimum depth, % cratonic and % discontinuous. We then evaluated relative support for a model including only these predictors and the intercept, these predictors and the intercept plus predicted extinctions and survivors in a greenhouse–icehouse transition, and these predictors and the intercept plus predicted extinctions and survivors in an icehouse–greenhouse transition. We used a classification tree to illustrate how the predictors in the best-supported model partition Katian 5 genera into extinctions and survivors.

3. Results and discussion

The performance of GBM models in accurately classifying genera as extinctions or survivors varied across intervals (figure 1), with the Katian 1, Katian 3 and Hirnantian models being notably poor. The marginal effects of predictors on extinction risk in each interval, and their relative influence on predicting extinction risk/survival, are shown as partial dependence plots in figure 1. Relative influence of predictors varied among intervals, but compared with other timeslices, the timeslice coinciding with the major pulse of the LOME (Katian 5) stands out for the unusual relative influence and marginal effects of palaeolatitude range, minimum depth, % discontinuous and % cratonic. Great circle distance is also an

important predictor of extinction risk during Katian 5, but similar relationships between great circle distance and marginal risk are apparent in several other timeslices. In contrast, although palaeolatitude range is an important predictor of extinction risk in some other timeslices, no other timeslices show a similar relationship between palaeolatitude range and marginal risk: genera with extremely restricted palaeolatitude ranges (often those restricted to a single locality or region) exhibit elevated marginal risk in some timeslices, but only Katian 5 exhibits a broad plateau of elevated marginal risk among genera with palaeolatitude ranges less than about 35°.

Katian 5 brachiopod extinctions are also strikingly selective with respect to bathymetric distribution. Genera that ranged into shallower waters (BA 1–2) experienced much lower extinction rates than those restricted to deeper waters (BA 3–6). The pattern is not limited to a single region but is apparent across a broad palaeolatitude range (figure 2), implying that it reflects the operation of a global-scale environmental driver and that the previously noted demise of the widespread deep water (BA 5–6) *Foliomena* fauna [14,22] is only the most extreme manifestation of a broader selective sweep. Interpretation of this pattern is complicated by uncertainty regarding the actual depth ranges represented by BAs (which are restricted to shelf and slope settings) [26], but the sign of the depth signal is informative. Cooling alone would be expected to steepen bathymetric temperature profiles and preferentially cause the extinction of genera restricted to warmer surface waters rather than those restricted to cooler deeper water. It is more likely, therefore, that the bathymetric extinction gradient reflects some other change in water mass characteristics.

Several lines of evidence, including biomarkers [37], nitrogen isotopes [38,39], molybdenum isotopes [14,40], iron speciation [14] and black shale distributions [39], suggest that the onset of the Hirnantian icehouse climate state was accompanied by increased oxygenation of shelf environments. Many of the species found in deeper-water environments during the late Katian were small and thin-shelled [41], like modern low-oxygen specialists [42]. Benthic species that had adapted to relatively low-oxygen conditions prevailing in deeper waters during the Katian greenhouse climate state may have been driven extinct by expansion of more oxygenated waters with their incumbent shallow-water faunas. This scenario is consistent with the contemporaneous extinction of multiple graptolite lineages thought to have inhabited denitrifying waters on the margins of oxygen minimum zones [16,39,43,44], including many older and previously extinction-resistant lineages [45]. An alternative interpretation is that the extinction of deeper-water genera reflects expansion of oxygen minimum zones and associated sulfidic conditions [14], but recent studies [46] do not support this hypothesis. Changes in oxygenation would not necessarily affect all taxa similarly, and whether such changes can explain other selective patterns such as the disproportionate extinction of trilobites with planktonic life stages [17] is not clear.

Both the first (end Katian) and second (late Hirnantian) pulses of the LOME exhibit slightly elevated extinction risk for genera occurring primarily in cratonic seaways relative to those occurring primarily in open-shelf settings (figure 1). Although this effect is weak, it is notable because it is opposite to that observed in most other intervals, during which genera restricted to open-shelf environments exhibit slightly greater

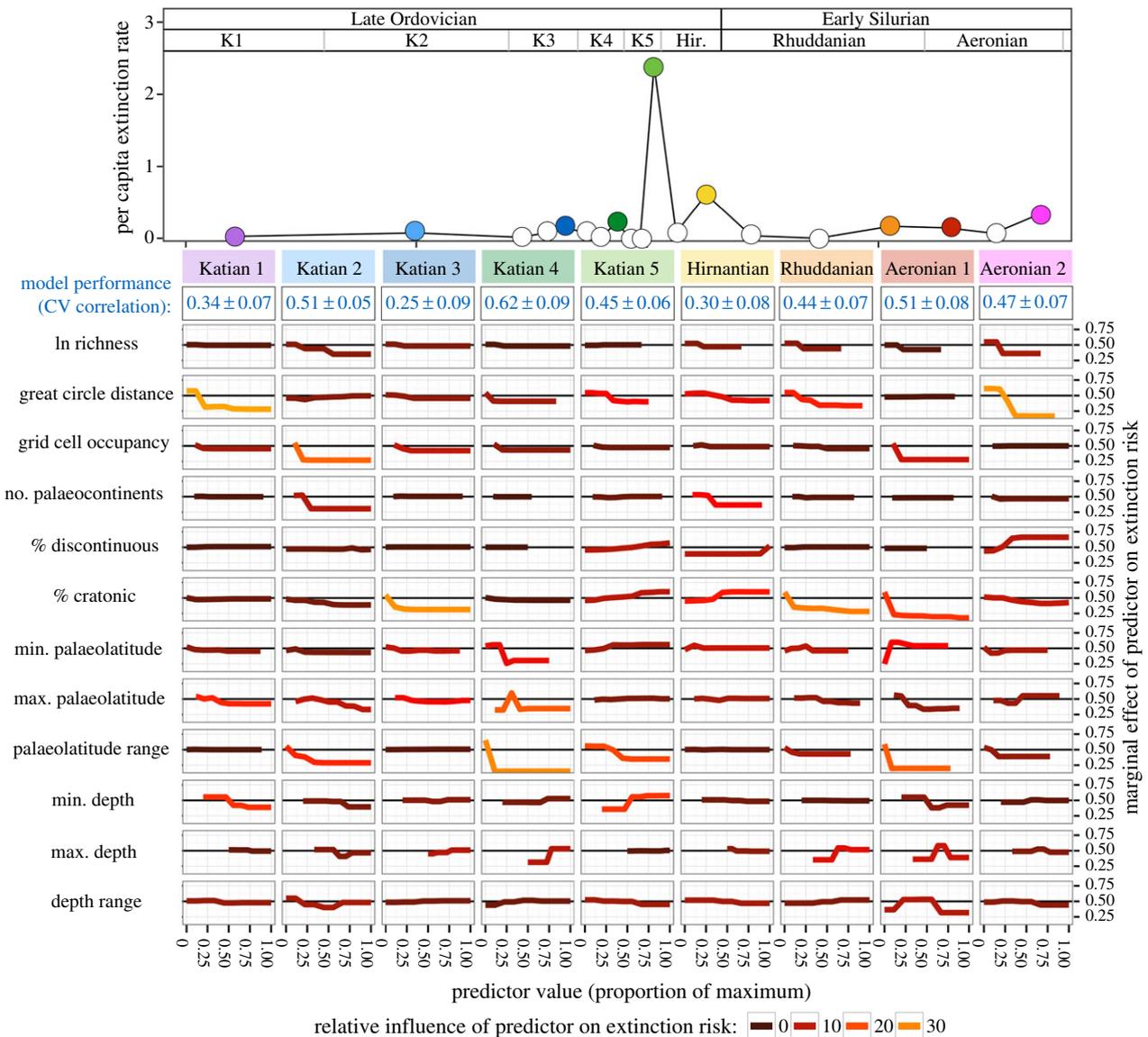


Figure 1. Top: per capita extinction rates of brachiopod genera, in lineage million years, during each of 19 Late Ordovician–Early Silurian timeslices. Coloured points indicate the timeslices with at least 10 extinctions analysed in, bottom panel. Bottom: partial dependence plots for GBM models trained on extinction patterns in each interval. Values above 0.5 indicate a tendency for genera with the given predictor value to go extinct when all other variables are held constant, values below 0.5 indicate a tendency for genera with the given predictor value to survive. The total range of marginal effects is independently scaled within each interval in order to highlight the effects of each predictor on relative extinction risk. x -axis units are rescaled to the maximum value for each predictor. Line colours reflect the relative influence of each predictor in each interval. Mean and s.e. of correlations from 20-fold cross-validation of model predictions on holdout observations (40%) are given in blue text below timeslice names. Untransformed predictor ranges are 1–57 species (ln richness), 10–12 000 km (great circle distance), 1–100% occupancy (grid cell occupancy), 1–11 palaeocontinents/terranes (no. alaeocontinents), 0–100% of localities (% cratonic, % discontinuous), 0–90° absolute latitude (maximum and minimum palaeolatitude, palaeolatitude range) and brachiopod depth assemblage 1–6 (minimum and maximum depth, depth range).

risk than cratonic genera. Preferential extinction of cratonic genera contrasts with preferential survival of such taxa during Meso-Cenozoic mass extinctions [47], but is consistent with previous suggestions that some latest Katian extinctions were driven by eustatically forced regression and draining of cratonic seaways [7,48]. It should be noted, however, that % discontinuous is also weakly predictive of extinction risk in both Katian 5 and (to a lesser extent) the Hirnantian. Moreover, in Katian 5, the marginal risks associated with % discontinuous and % cratonic are rather similar (figure 1).

The positive association between % discontinuous and extinction risk, though weak, raises the possibility that some proportion of the extinctions that appear to occur at the Katian–Hirnantian boundary represent artificial range truncations resulting from reduced preservation probability. Such truncations would not be expected to affect all genera

equally. The draining of cratonic seaways may have led to genuine extinctions of genera with dominantly cratonic distributions, but these genera might also be expected to have experienced disproportionate decline in preservation potential. In shelf areas most preserved sequences record substantial shoaling [3,6,19,49,50], raising the possibility that deeper-water genera may also have experienced a disproportionate decline in preservation potential owing to reduced sampling of appropriate environments.

We evaluated the likelihood that observed extinction selectivity patterns are driven by stratigraphic range truncations by comparing mean predictor values in genera that appear to go extinct in Katian 5, ‘range-through’ genera that survive but have a post-Katian 5 sampling gap, and genera that survive and occur in the subsequent (early Hirnantian) timeslice. In the extreme case that all apparent extinctions actually represented stratigraphic range truncations, the factors

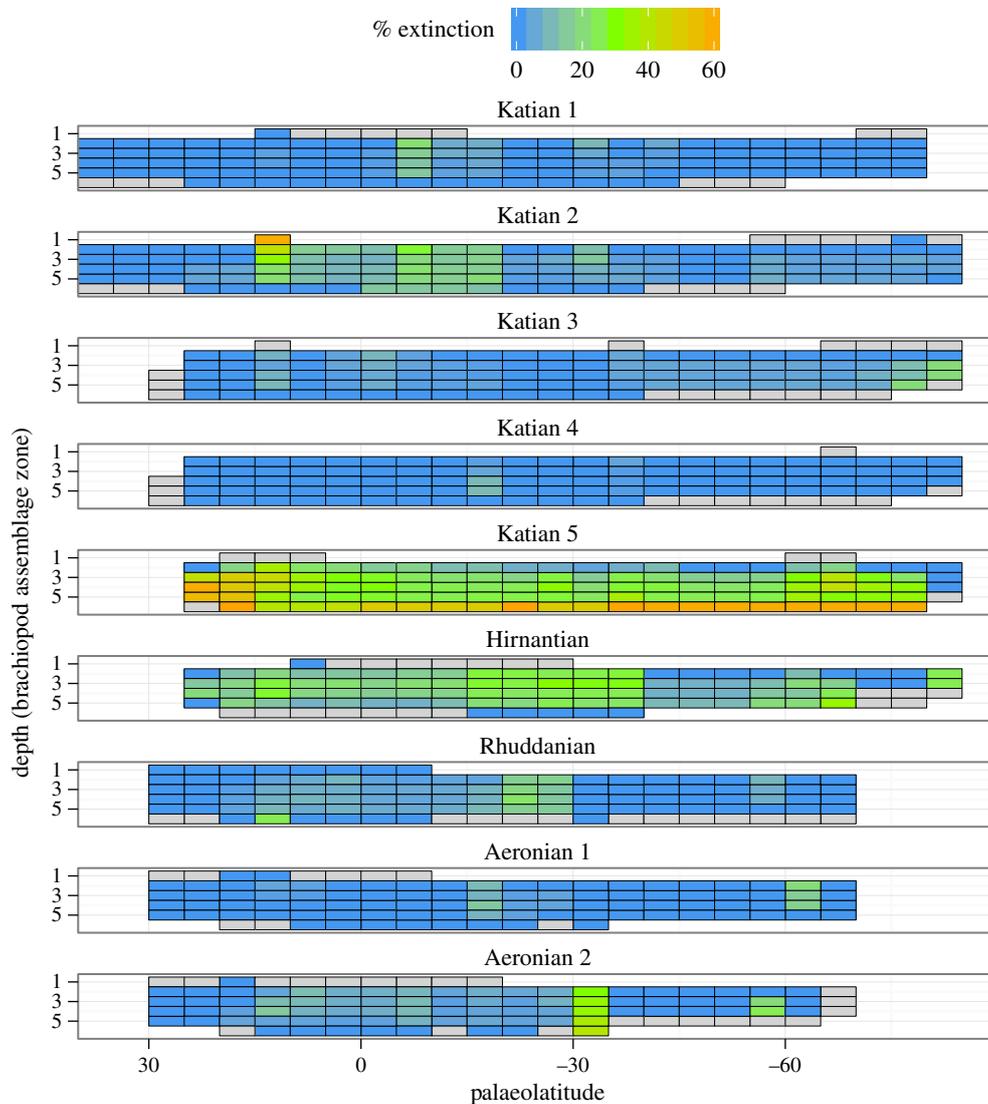


Figure 2. Percentage extinction of genera inhabiting different palaeolatitudinal and depth bins during 10 Late Ordovician–Early Silurian intervals (oldest at top). Genera were assumed to be present throughout their recorded palaeolatitudinal and depth ranges. Depth ranges are based on brachiopod depth assemblage zones ranging from 1 (shallowest) to 6 (deepest). Only palaeolatitudinal depth cells occupied by at least one genus are outlined; those occupied by fewer than three genera are indicated by grey fill. The first (Katian 5) pulse of the Late Ordovician mass extinction is distinguished by the overall intensity of extinction, but also by the strongly selective extinction of deeper-water genera.

important in predicting extinction risk would instead be interpretable as predictors of going unsampled in the early Hirnantian, and we would expect range-through genera and ‘extinct’ genera to exhibit similar mean predictor values. Of the four Katian 5 predictors included in at least 75% of the top multiple logistic regression models, % discontinuous and (to a lesser extent) % cratonic exhibit such a pattern (electronic supplementary material, figure S2). Mean palaeolatitudinal range and minimum depth values of range-through genera are intermediate between those of extinct genera and genera that survive with no sampling gap. This suggests that, although apparent extinctions in Katian 5 may represent a mixture of genuine extinctions and range truncations, observed palaeolatitudinal and bathymetric extinction selectivity patterns are unlikely to be artefacts of record bias.

The unusual importance of palaeolatitudinal range in the latest Katian implies that changing sea surface temperatures played a major role in driving extinctions. Latitudinal ranges are closely linked to thermal tolerance ranges among extant aquatic ectotherms [51–53], and sea surface temperatures are the single most important determinant of biogeographic

structure in the coastal oceans [54,55]. The thermal tolerance ranges of some extant marine invertebrate species have been conserved on million-year time scales [56], and many species shifted their ranges in response to Cenozoic and Quaternary climate changes [55,57,58].

Preferential survival of genera with wide palaeolatitudinal ranges does not necessarily implicate cooling; broad thermal tolerance range would be expected to buffer against extinction during any rapid climate change, and the Latest Ordovician world likely experienced glacial–interglacial oscillations analogous to those of the Cenozoic. It has been argued that the first pulse of the LOME corresponds not to the greenhouse–icehouse transition but rather to the first interglacial within the icehouse state [19], and the selective extinction of narrow-ranging genera is also consistent with this hypothesis. However, a multivariate logistic regression model that includes the four selected predictors plus predictions about which genera would be expected to go extinct owing to habitat loss during a greenhouse–icehouse transition strongly outperforms both the model including only the four original predictors and the model including predictions about which

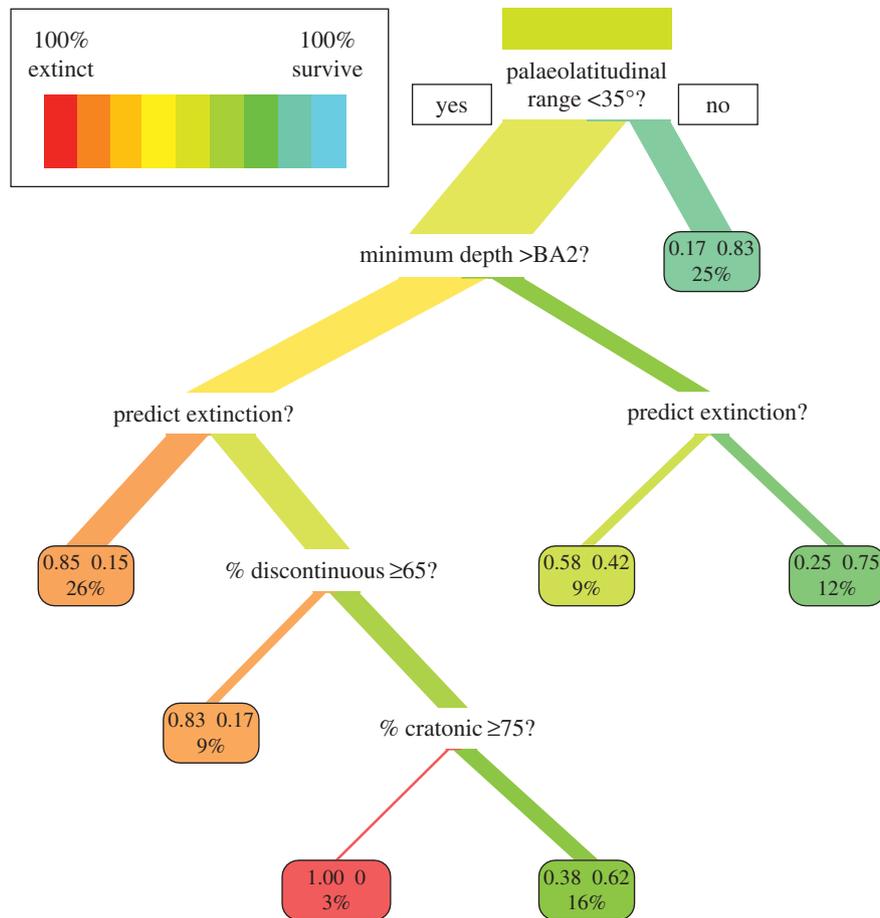


Figure 3. Classification tree shows partitioning of Katian 5 genera into extinctions and survivors based on the four selected predictors and predicted extinction during a greenhouse–icehouse transition. Branch thicknesses are scaled to the number of genera included in each split. Each node included the proportion of genera in the node that go extinct (top left), the proportion that survive (top right) and the percentage of all 205 Katian 5 genera that are included in the node (bottom). Colour scale indicates the percentage of extinctions versus survivors in each node.

Table 1. Comparison of three different multiple logistic regression models using the modified Akaike information criterion (AICc): ‘no prediction’ (extinction \sim palaeolatitudinal range + minimum depth + % cratonic + % discontinuous), ‘greenhouse–icehouse prediction’ (extinction \sim palaeolatitudinal range + minimum depth + % cratonic + % discontinuous + predicted extinction/survival in a greenhouse–icehouse transition) and ‘icehouse–greenhouse prediction’ (extinction \sim palaeolatitudinal range + minimum depth + % cratonic + % discontinuous + predicted extinction/survival in an icehouse–greenhouse transition). The ‘greenhouse–icehouse prediction’ model, in which the expected survival or extinction of a genus in an icehouse climate state is predicted based on its distribution in an assumed greenhouse climate state, is favoured over the other models by evidence ratios $>10:1$. K = number of estimated parameters for each model (including intercept), $\Delta AICc$ = AICc difference between model and ‘best’ model, AICc Wt = proportional support for model, LL = log likelihood.

model name	K	AIC _c	$\Delta AICc$	AIC _c Wt	LL
greenhouse–icehouse prediction	6	221.06	0	0.99	−104.32
no prediction	5	230.75	9.68	0.01	−110.22
icehouse–greenhouse prediction	6	232.51	11.45	0.00	−110.05

genera would be expected to go extinct in a hypothetical icehouse–greenhouse transition (table 1). Consequently, we conclude that cooling and equatorward shift of climate zones led to the extinction of many species (and thus genera) that had narrow thermal tolerance ranges and inhabited coastlines or cratonic seaways of limited latitudinal extent.

Extinction selectivity patterns during Katian 5 can be visualized with a classification tree (figure 3). Unsurprisingly, given that our predictions for extinction versus survival in a greenhouse–icehouse transition are necessarily crude estimates, palaeolatitudinal range remains an important predictor of extinction risk even in models that include these predictions. Only 17% of genera with palaeolatitudinal ranges more than

35° go extinct (figure 3), but of those with ranges less than 35° , extinction is substantially higher in genera confined to depths of BA3 or deeper than in genera that ranged into shallower waters. Predicted extinction in a greenhouse–icehouse transition is selected as a further split in both the exclusively deep water and shallower subsets of the narrow-ranging genera, with further splits on % cratonic and % discontinuous among narrow-ranging deep-water genera that are not otherwise predicted to go extinct (figure 3). This analysis thus confirms that aspects of thermal tolerance range and bathymetric distribution seem to have been more important than occupying cratonic seaways and/or regions with widespread stratigraphic truncations in determining which genera went extinct and which survived.

The poor performance of the Hirnantian GBM model (figure 1) indicates that none of the variables considered here are particularly effective predictors of extinction risk during the smaller second pulse of the LOME in the mid- to late Hirnantian. A previous analysis of the Laurentian record also failed to find effective predictors of extinction risk for the second LOME pulse [13]. It has been suggested that extinctions during this interval are related to warming and shoaling of oxygen-poor waters during partial deglaciation [14,39], but we observe no strong depth or palaeolatitudinal signal in the distribution of extinction risk. Future analyses incorporating additional environmental factors (e.g. the percentage of sites occupied by a given genus in the late Hirnantian that are overlain by laminated black shales in the Rhuddanian) may be able to resolve a clearer selective signal.

Our results suggest an explanation for one of the enduring questions about the LOME: why did the Late Ordovician greenhouse–icehouse transition cause a major mass extinction when subsequent greenhouse–icehouse transitions did not? In fact, both the late Eocene–Oligocene transition and the Pliocene–Pleistocene descent into full icehouse conditions were associated with significant extinction pulses [59–63], but these pulses were predominantly focused on tropical and subtropical regions and were not of comparable magnitude. The contrast may reflect three important differences in starting state between Late Ordovician and Caenozoic greenhouse–icehouse transitions.

(1) Early Palaeozoic dissolved oxygen levels may have been substantially below typical modern levels, especially in outer shelf and slope settings [64]. Whereas the Late Ordovician greenhouse–icehouse transition seems to have been accompanied by a substantial increase in shelf oxygenation [37–39,43], there is little evidence for a comparable increase during the Caenozoic transition. There are many potential explanations for this contrast, but one key factor may be Phanerozoic evolutionary trends that have acted to deepen the e-folding depth of organic remineralization [65]. Such a shift would deepen oxygen minimum zones and reduce the exposure of shallow benthic habitats to substantial fluctuations in dissolved oxygen content.

(2) The late Katian world was an extreme highstand with extensive flooding of continents, but cratonic seaways were of limited extent in the Caenozoic and almost entirely gone by the Pliocene [66] (electronic supplementary material, figure S3). Loss of cratonic faunas, many of which probably experienced substantial oxygen fluctuations [67] and would have had difficulty dispersing into open-shelf settings [68], was almost certainly a more important driver of extinctions

during Late Ordovician cooling and sea level fall than during Caenozoic cooling episodes.

(3) Except for the supercontinent of Gondwana, the Late Ordovician world was characterized by widely dispersed island continents and an abundance of isolated terranes [24,33], whereas the Caenozoic world was generally characterized by extensive continuous north–south coastlines [69] (electronic supplementary material, figure S3). Caenozoic continental configurations would thus have allowed many marine species to shift their ranges equatorward in response to shifting climate zones without having to disperse across open ocean basins. The Late Ordovician configuration may have had a greater tendency to ‘trap’ species on coastlines from which they could not easily shift their ranges equatorward, accounting for the selective extinction of genera with narrow palaeolatitudinal ranges.

Our finding may also help to explain the observation that the LOME was taxonomically relatively non-selective and seems to have had far less long-term impact on the taxonomic and ecological structure of marine ecosystems than other major mass extinction events [70,71]. Whereas both the Permian–Triassic and Cretaceous–Palaeogene events removed groups that had previously been diverse and ecologically dominant, very few higher taxa disappeared during the LOME. Our analyses show that among rhynchonelliform brachiopods extinction was in fact strongly selective, but along bathymetric and biogeographic gradients that would have affected most major taxa. Consequently, although taxonomic losses were very high, these losses were relatively evenly distributed across ecospace. With the re-establishment of background environmental conditions surviving lineages rapidly diversified [72] into similar regions of ecospace, giving rise to ecosystems structurally similar to those that preceded the LOME.

Data accessibility. Data used in the analyses reported here are deposited at Dryad: <http://dx.doi.org/10.5061/dryad.66vb3>. The data can also be downloaded, along with R scripts to reproduce all analyses, at GitHub: <https://github.com/sethfinnegan/Ord.Brach.Extinctions>. Code.git. See the electronic supplementary material for further instructions.

Author contributions. All authors jointly conceived of the study. S.F. processed the data, conducted all statistical analyses and drafted the manuscript and most figures; C.M.Ø.R. and D.A.T.H. compiled the database and produced some supplementary figures. All authors gave final approval for publication.

Competing interests. We declare we have no competing interests.

Funding. The authors acknowledge generous support from the David and Lucile Packard Foundation, the Danish Council for Independent Research and the Villum Foundation Young Investigator Programme.

References

- Bambach RK. 2006 Phanerozoic biodiversity mass extinctions. *Annu. Rev. Earth Planet. Sci.* **34**, 127–155. (doi:10.1146/annurev.earth.33.092203.122654)
- Raup DM, Sepkoski JJ, Stigler SM. 1983 Mass extinctions in the fossil record. *Science* **219**, 1240–1241. (doi:10.1126/science.219.4589.1240)
- Sheehan PM. 2001 The Late Ordovician mass extinction. *Annu. Rev. Earth Planet. Sci.* **29**, 331–364. (doi:10.1146/annurev.earth.29.1.331)
- Brenchley PJ, Carden GA, Hints L, Kaljo D, Marshall JD, Martma T, Meidla T, Nolvak J. 2003 High-resolution stable isotope stratigraphy of Upper Ordovician sequences: constraints on the timing of bioevents and environmental changes associated with mass extinction and glaciation. *Geol. Soc. Am. Bull.* **115**, 89–104. (doi:10.1130/0016-7606(2003)115<0089:HRSISO>2.0.CO;2)
- Harper DAT, Hammarlund EU, Rasmussen CMØ. 2014 End Ordovician extinctions: a coincidence of causes. *Gondwana Res.* **25**, 1294–1307. (doi:10.1016/j.gr.2012.12.021)
- Brenchley PJ, Marshall JD, Carden GAF, Robertson DBR, Long DGF, Meidla T, Hints L, Anderson TF. 1994 Bathymetric and isotopic evidence for the short-lived Late Ordovician glaciation in a greenhouse period. *Geology* **22**, 295–298. (doi:10.1130/G01000a.1)

- 1130/0091-7613(1994)022<0295:BAIEFA>2.3.CO;2)
7. Sheehan PM. 1973 The relation of Late Ordovician glaciation to the Ordovician–Silurian changeover in North American brachiopod faunas. *Lethaia* **6**, 147–154. (doi:10.1111/j.1502-3931.1973.tb01188.x)
 8. Brechley PJ. 1984 Late Ordovician extinctions and their relationship to the Gondwana glaciation. In *Fossils and climate* (ed. PJ Brechley), pp. 291–327. New York, NY: John Wiley and Sons, Inc.
 9. Finnegan S, Bergmann K, Eiler JM, Jones DS, Fike DA, Eisenman I, Hughes NC, Tripathi AK, Fischer WW. 2011 The magnitude and duration of Late Ordovician–Early Silurian glaciation. *Science* **331**, 903–906. (doi:10.1126/science.1200803)
 10. Barnes CR. 1986 The faunal extinction event near the Ordovician–Silurian boundary: a climatically induced crisis. In *Global bio-events* (ed. OH Walliser), pp. 119–126. Berlin, Germany: Springer.
 11. Jablonski D. 1991 Extinctions; a paleontological perspective. *Science* **253**, 754–757. (doi:10.1126/science.253.5021.754)
 12. Vandenbroucke TRA *et al.* 2010 Polar front shift and atmospheric CO₂ during the glacial maximum of the Early Paleozoic Icehouse. *Proc. Natl Acad. Sci. USA* **107**, 14 983–14 986. (doi:10.1073/pnas.1003220107)
 13. Finnegan S, Heim NA, Peters SE, Fischer WW. 2012 Climate change and the selective signature of the Late Ordovician mass extinction. *Proc. Natl Acad. Sci. USA* **109**, 6829–6834. (doi:10.1073/pnas.1117039109)
 14. Hammarlund EU *et al.* 2012 A sulfidic driver for the end-Ordovician mass extinction. *Earth Planet. Sci. Lett.* **331–332**, 128–139. (doi:10.1016/j.epsl.2012.02.024)
 15. Finney SC, Berry WBN, Cooper JD, Ripperdan RL, Sweet WC, Jacobson SR, Soufiane A, Achab A, Noble PJ. 1999 Late Ordovician mass extinction; a new perspective from stratigraphic sections in central Nevada. *Geology* **27**, 215–218. (doi:10.1130/0091-7613(1999)027<0215:L0MEAN>2.3.CO;2)
 16. Xu C, Melchin MJ, Sheets HD, Mitchell CE, Jun-Xuan F. 2005 Patterns and processes of latest Ordovician graptolite extinction and recovery based on data from South China. *J. Paleontol.* **79**, 842–861. (doi:10.1666/0022-3360(2005)079[0842:PAPOLO]2.0.CO;2)
 17. Chatterton BDE, Speyer SE. 1989 Larval ecology, life history strategies, and patterns of extinction and survivorship among Ordovician Trilobites. *Paleobiology* **15**, 118–132.
 18. Harper DAT, Jiayu R. 2008 Completeness of the Hirnantian brachiopod record: spatial heterogeneity through the end Ordovician extinction event. *Lethaia* **41**, 195–197. (doi:10.1111/j.1502-3931.2008.00098.x)
 19. Ghienne J-F *et al.* 2014 A Cenozoic-style scenario for the end-Ordovician glaciation. *Nat Commun* **5**, 4485.
 20. Holland SM, Patzkowsky ME. 2015 The stratigraphy of mass extinction. *Paleontology* **58**, 903–924. (doi:10.1111/pala.12188)
 21. Rong J, Boucot AJ, Harper DAT, Zhan R, Neuman RB. 2006 Global analyses of brachiopod faunas through the Ordovician and Silurian transition: reducing the role of the Lazarus effect. *Can. J. Earth Sci.* **43**, 23–39. (doi:10.1139/e05-089)
 22. Rasmussen CMØ, Harper DAT. 2011 Interrogation of distributional data for the End Ordovician crisis interval: where did disaster strike? *Geol. J.* **46**, 478–500. (doi:10.1002/gj.1310)
 23. Rasmussen CMØ. 2014 Phylogeography of Ordovician–Silurian rhynchonelliformean brachiopods: tracking higher order distributional patterns, radiations and extinctions in 4D. *GFF* **136**, 223–228. (doi:10.1080/11035897.2013.847485)
 24. Rasmussen CMØ, Harper DAT. 2011 Did the amalgamation of continents drive the end Ordovician mass extinctions? *Palaogeogr. Palaeoclimatol. Palaeoecol.* **311**, 48–62. (doi:10.1016/j.palaeo.2011.07.029)
 25. Bergström SM, Chen X, Gutiérrez-Marco JC, Dronov A. 2009 The new chronostratigraphic classification of the Ordovician System and its relations to major regional series and stages and to $\delta^{13}\text{C}$ chemostratigraphy. *Lethaia* **42**, 97–107. (doi:10.1111/j.1502-3931.2008.00136.x)
 26. Brett CE, Boucot AJ, Jones B. 1993 Absolute depths of Silurian benthic assemblages. *Lethaia* **26**, 25–40. (doi:10.1111/j.1502-3931.1993.tb01507.x)
 27. Boucot AJ. 1975 *Evolution and extinction rate controls*. New York, NY: Elsevier.
 28. Brechley PJ, Harper DAT. 1998 *Palaeoecology: ecosystems, environments and evolution*. London, UK: Chapman and Hall.
 29. R Core Team. 2015 *R: a language and environment for statistical computing*. Vienna, Austria: R Foundation for Statistical Computing.
 30. Jablonski D. 1996 Causes and consequences of mass extinctions: a comparative approach. In *Dynamics of extinction* (ed. DK Elliott), pp. 183–229. New York, NY: John Wiley and Sons.
 31. Herrmann AD, Haupt BJ, Patzkowsky ME, Seidov D, Slingerland RL. 2004 Response of Late Ordovician paleoceanography to changes in sea level, continental drift, and atmospheric pCO₂; potential causes for long-term cooling and glaciation. *Palaogeogr. Palaeoclimatol. Palaeoecol.* **210**, 385–401. (doi:10.1016/j.palaeo.2004.02.034)
 32. Cocks LRM, Torsvik TH. 2011 The Palaeozoic geography of Laurentia and western Laurussia: a stable craton with mobile margins. *Earth Sci. Rev.* **106**, 1–51. (doi:10.1016/j.earscirev.2011.01.007)
 33. Cocks LRM, Torsvik TH. 2002 Earth geography from 500 to 400 million years ago: a faunal and palaeomagnetic review. *J. Geol. Soc.* **159**, 631–644. (doi:10.1144/0016-764901-118)
 34. Elith J, Leathwick JR, Hastie T. 2008 A working guide to boosted regression trees. *J. Anim. Ecol.* **77**, 802–813. (doi:10.1111/j.1365-2656.2008.01390.x)
 35. Hijmans RJ, Phillips S, Elith JL. 2015 *dismo: species distribution modeling* (cited 20 December 2015).
 36. Hosmer DW, Lemeshow S. 2000 *Applied logistic regression*. New York, NY: Wiley.
 37. Rohrsen M, Love GD, Fischer W, Finnegan S, Fike DA. 2012 Lipid biomarkers record fundamental changes in the microbial community structure of tropical seas during the Late Ordovician Hirnantian glaciation. *Geology* **41**, 127. (doi:10.1130/G33671.1)
 38. Luo G, Algeo TJ, Zhan R, Yan D, Huang J, Liu J, Xie S. 2016 Perturbation of the marine nitrogen cycle during the Late Ordovician glaciation and mass extinction. *Palaogeogr. Palaeoclimatol. Palaeoecol.* **448**, 339–348. (doi:10.1016/j.palaeo.2015.07.018)
 39. Melchin MJ, Mitchell CE, Holmden C, Štorch P. 2013 Environmental changes in the Late Ordovician–early Silurian: review and new insights from black shales and nitrogen isotopes. *Geol. Soc. Am. Bull.* **125**, 1635 (doi:10.1130/B30812.1)
 40. Zhou L, Algeo TJ, Shen J, Hu Z, Gong H, Xie S, Huang J, Gao S. 2015 Changes in marine productivity and redox conditions during the Late Ordovician Hirnantian glaciation. *Palaogeogr. Palaeoclimatol. Palaeoecol.* **420**, 223–234. (doi:10.1016/j.palaeo.2014.12.012)
 41. Zhan R-B, Jin J. 2005 New data on the Foliomena Fauna (Brachiopoda) from the Upper Ordovician of South China. *J. Paleontol.* **79**, 670–686. (doi:10.1666/0022-3360(2005)079[0670:NDOTFF]2.0.CO;2)
 42. Levin LA. 2003 Oxygen minimum zone benthos: adaptation and community response to hypoxia. *Oceanogr. Mar. Biol.* **41**, 1–45.
 43. Finney SC, Berry WBN, Cooper JD. 2007 The influence of denitrifying seawater on graptolite extinction and diversification during the Hirnantian (latest Ordovician) mass extinction event. *Lethaia* **40**, 281–291. (doi:10.1111/j.1502-3931.2007.00027.x)
 44. Berry WB, Wilde P, Quinby-Hunt MS. 1987 The oceanic non-sulfidic oxygen minimum zone: a habitat for graptolites. *Bull. Geol. Soc. Den.* **35**, 103–114.
 45. Crampton JS, Cooper RA, Sadler PM, Foote M. 2016 Greenhouse–icehouse transition in the Late Ordovician marks a step change in extinction regime in the marine plankton. *Proc. Natl Acad. Sci. USA* **113**, 1498–1503. (doi:10.1073/pnas.1519092113)
 46. Jones DS, Fike DA. 2013 Dynamic sulfur and carbon cycling through the end-Ordovician extinction revealed by paired sulfate–pyrite $\delta^{34}\text{S}$. *Earth Planet. Sci. Lett.* **363**, 144–155. (doi:10.1016/j.epsl.2012.12.015)
 47. Miller AI, Foote M. 2009 Epicontinental seas versus open-ocean settings: the kinetics of mass extinction and origination. *Science* **326**, 1106–1109. (doi:10.1126/science.1180061)
 48. Sheehan PM. 1988 Late Ordovician events and the terminal Ordovician extinction. *N. M. Bur. Mines Miner. Resour. Mem.* **44**, 405–415.
 49. Arthur MA, Gibbs MT, Kump LR, Patzkowsky ME, Pinkus DS, Sheehan PM. 1999 A weathering hypothesis for glaciation at high atmospheric pCO₂ during the Late Ordovician. *Palaogeogr. Palaeoclimatol. Palaeoecol.* **152**, 173–187. (doi:10.1016/S0031-0182(99)00046-2)
 50. Harris MT, Sheehan PM, Ainsaar L, Hints L, Männik P, Nõlvak J, Rubel M. 2004 Upper Ordovician sequences of western Estonia. *Palaogeogr.*

- Palaeoclimatol. Palaeoecol.* **210**, 135–148. (doi:10.1016/j.palaeo.2004.02.045)
51. Sunday JM, Bates AE, Dulvy NK. 2012 Thermal tolerance and the global redistribution of animals. *Nat. Clim. Change* **2**, 686–690. (doi:10.1038/nclimate1539)
 52. Sunday JM, Bates AE, Dulvy NK. 2010 Global analysis of thermal tolerance and latitude in ectotherms. *Proc. R. Soc. B* **278**, 1823–1830. (doi:10.1098/rspb.2010.1295)
 53. Calosi P, Bilton DT, Spicer JL, Votier SC, Atfield A. 2010 What determines a species' geographical range? Thermal biology and latitudinal range size relationships in European diving beetles (Coleoptera: Dytiscidae). *J. Anim. Ecol.* **79**, 194–204. (doi:10.1111/j.1365-2656.2009.01611.x)
 54. Belanger CL, Jablonski D, Roy K, Berke SK, Krug AZ, Valentine JW. 2012 Global environmental predictors of benthic marine biogeographic structure. *Proc. Natl Acad. Sci. USA* **109**, 14 046–14 051. (doi:10.1073/pnas.1212381109)
 55. Valentine JW. 1973 *Evolutionary paleoecology of the marine biosphere*, 1st edn. Englewood Cliffs, NJ: Prentice-Hall.
 56. Saupe EE, Hendricks JR, Portell RW, Dowsett HJ, Haywood A, Hunter SJ, Lieberman BS. 2014 Macroevolutionary consequences of profound climate change on niche evolution in marine molluscs over the past three million years. *Proc. R. Soc. B* **281**, 20141995. (doi:10.1098/rspb.2014.1995)
 57. Hall CA. 2002 *Nearshore marine paleoclimatic regions, increasing zoogeographic provinciality, molluscan extinctions, and paleoshorelines, California: Late Oligocene (27 Ma) to Late Pliocene (2.5 Ma)*. Boulder, CO: Geological Society of America.
 58. Roy K, Jablonski D, Valentine JW. 1995 Thermally anomalous assemblages revisited: patterns in the extraprovincial latitudinal range shifts of pleistocene marine mollusks. *Geology* **23**, 1071–1074. (doi:10.1130/0091-7613(1995)023<1071:TAARPI>2.3.CO;2)
 59. Hansen TA. 1987 Extinction of late eocene to oligocene molluscs: relationship to shelf area, temperature changes, and impact events. *PALAIOS* **2**, 69–75. (doi:10.2307/3514573)
 60. Raffi S, Stanley SM, Marasti R. 1985 Biogeographic Patterns and Plio-Pleistocene Extinction of Bivalvia in the Mediterranean and Southern North Sea. *Paleobiology* **11**, 368–388.
 61. Stanley SM. 1986 Anatomy of a regional mass extinction: plio-pleistocene decimation of the Western Atlantic Bivalve Fauna. *PALAIOS* **1**, 17–36. (doi:10.2307/3514456)
 62. Pearson PN, McMillan IK, Wade BS, Jones TD, Coxall HK, Bown PR, Lear CH. 2008 Extinction and environmental change across the Eocene–Oligocene boundary in Tanzania. *Geology* **36**, 179–182. (doi:10.1130/G24308A.1)
 63. Ivany LC, Patterson WP, Lohmann KC. 2000 Cooler winters as a possible cause of mass extinctions at the Eocene/Oligocene boundary. *Nature* **407**, 887–890. (doi:10.1038/35038044)
 64. Sperling EA, Wolock CJ, Morgan AS, Gill BC, Kunzmann M, Halverson GP, Macdonald FA, Knoll AH, Johnston DT. 2015 Statistical analysis of iron geochemical data suggests limited late Proterozoic oxygenation. *Nature* **523**, 451–454. (doi:10.1038/nature14589)
 65. Meyer KM, Ridgwell A, Payne JL. 2016 The influence of the biological pump on ocean chemistry: implications for long-term trends in marine redox chemistry, the global carbon cycle, and marine animal ecosystems. *Geobiology* **14**, 207–219. (doi:10.1111/gbi.12176)
 66. Scotese CR. 2001 *Atlas of earth history, volume 1: paleogeography*. Arlington, TX: PALEOMAP Project.
 67. Peters SE. 2007 The problem with the Paleozoic. *Paleobiology* **33**, 165–181. (doi:10.1666/06067.1)
 68. Stanley SM. 2010 Thermal barriers and the fate of perched faunas. *Geology* **38**, 31–34. (doi:10.1130/G30295.1)
 69. Valentine JW, Moores EM. 1970 Plate-tectonic regulation of faunal diversity and sea level: a model. *Nature* **228**, 657–659. (doi:10.1038/228657a0)
 70. Droser ML, Bottjer DJ, Sheehan PM, McGhee GR. 2000 Decoupling of taxonomic and ecologic severity of Phanerozoic marine mass extinctions. *Geology* **28**, 675–678. (doi:10.1130/0091-7613(2000)28<675:DOTAES>2.0.CO;2)
 71. McGhee GR, Sheehan PM, Bottjer DJ, Droser ML. 2004 Ecological ranking of Phanerozoic biodiversity crises: ecological and taxonomic severities are decoupled. *Palaeogeogr. Palaeoclimatol. Palaeoecol.* **211**, 289–297. (doi:10.1016/j.palaeo.2004.05.010)
 72. Krug AZ, Patzkowsky ME. 2004 Rapid recovery from the Late Ordovician mass extinction. *Proc. Natl Acad. Sci. USA* **101**, 17 605–17 610. (doi:10.1073/pnas.0405199102)